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## TERPENOID AND OTHER CONSTITUENTS OF HERNANDIA VOYRONI AND ANTHOCLEISTA AMPLEXICAULIS\*

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**Key Word Index**—Hernandia voyroni, Hernandiaceae; Anthocleista amplexicaulis: Loganiaceae; terpenoids; (+)-perillaldehyde; swertiamarin; (+)-bornesitol

Plant sources. H. voyroni Jum. (Hernandiaceae), A. amplexicaulis (Loganiaceae). R. Pernet¹ has identified perillaldehyde as a constituent of the oil obtained from H. peltata Meissn. He also investigated extracts obtained from H. voyroni, whose alkaloid contents he estimated to be 0.5%, and in addition isolated 2.9% of an oil with camphor-like smell. We have further examined this oil and we have isolated (+)-perillaldehyde (1) from this species. Derivatives of 1 (oxime, semicarbazone) were prepared and compared with authenitic samples.

Swertiamarin (2) and (+)-bornesitol (3) were isolated from A. amplexicaulis, tetraacetyland dihydrotetraacetyl-2 were prepared, spectroscopically investigated and compared with authentic specimens. 2 has previously been isolated from A. procera.<sup>2</sup> 3 was transformed into its pentaacetyl derivative and the constitution of this product was elucidated by NMR spectroscopy. 3 has been previously found in Sarcocephalus diderichii (Rubiaceae).<sup>3</sup>

- \* Part 4 in the series "Plants from Madagascar"; for part 1, see SCHLITTLER, E. and WEBER, N. (1972) Lloydia 35, 181. Part 2, see SCHITTLER, E. and WEBER, N. (1972) Helv. Chim. Acta 55, 2061. Part 3, see WEBER, N. (1973) Chem. Ber. 106, 3769.
- <sup>1</sup> Pernet, R. (1971) Planta Medica 20, 314.
- <sup>2</sup> KOCH, M., PLAT, M., LEMEN, J. and JANOT, M. M. (1964) Bull. Soc. Chim. Fr. 403.
- <sup>3</sup> King, F. E. and Jurd, L. (1953) J. Chem. Soc. 1192.

## EXPERIMENTAL

M.ps are uncorrected. NMR spectra were recorded with TMS as an internal standard. UV in MeOH. IR in KBr-pellets. Optical rotation on a Perkin-Elmer-141 polarimeter. TLC were performed on neutral alumina type T (Merck), solvent CH<sub>2</sub>Cl<sub>2</sub>-MeOH (10:1). Column chromatography on alumina (Woelm, neutral). Bark and branches of H. voyroni were collected in November 1967 in Morondava/Madagascar, leaves of A. amplexicaulis in October 1967 in Perinet/Madagascar.

Extraction of H. voyroni. 100 g of stem bark and branches were extracted and worked up according to Schlittler et al.<sup>4</sup> Ethereal oils were separated by steam distillation and extracted from the aq. phase with Et<sub>2</sub>O. The ether residue yielded 0·3 g of crude oil which was purified by vacuum distillation. The main fraction (110-120°/12 mm) contained 1 and after redistillation a colourless oil with a camphor-like smell was obtained: UV  $\lambda_{\text{max}}$  228 (log 4·2), 307 (1·7) nm; IR (film)  $\nu_{\text{max}}$  2815 and 2723 (aldehyde), 1687 ( $\alpha$ , $\beta$ -unsat. aldehyde), 1645 (C=C) cm<sup>-1</sup> oxime: m.p. 100-101° (MeOH), [ $\alpha$ ]<sub>D</sub> + 147° ( $\alpha$  0·86; Et<sub>2</sub>O). compared (UV, IR, m.m.p.) with an authentic sample.

Semicarbazone. m.p. 195-197° (MeOH).

Extraction of A. amplexicaulis. 190 g ground leaves were defatted with petrol. and extracted with CHCl<sub>3</sub> and

EtOAc (5% H2O) according to Taylor-Smith.5

(a) Chloroform extract. Evaporation of the CHCl<sub>3</sub> extract left 6 g dark green residue which was chromatographed over 130 g alumina. The development of the chromatogram was followed up by TLC. Elution with acetone gave a dark green fraction (2·2 g, discarded); further elution with Me<sub>2</sub>CO-H<sub>2</sub>O (10:1) and Me<sub>2</sub>CO-H<sub>2</sub>O (1:1) afforded two fractions (1·8 and 1·0 g) which were pooled, dissolved in 10 ml MeOH and ppt. with 100 ml Et<sub>2</sub>O. The ppt, thus formed was dissolved in little MeOH, taken on an alumina column and eluted with MeOH-H<sub>2</sub>O (10:1). After evaporation of the solvent 1·0 g of a yellowish foam of 2 was obtained:  $[\alpha]_D = 105^{\circ}$  (c 0·97; MeOH).

Swertiamarin tetraacetate was obtained by acetylation of 2 (Ac<sub>2</sub>O pyridine at r.t. overnight), m.p. 194° (MeOH),  $[\alpha]_D = 116^\circ$  (c 1.06; CHCl<sub>3</sub>), compared (UV, IR, NMR) with authentic material (Found: C, 53·12; H, 5·44. C<sub>24</sub>H<sub>30</sub>O<sub>14</sub> requires: C, 53·13; H, 5·57%).

Dihydrosweriamarin tetraacetate was obtained by catalytic hydrogenation of 2 tetraacetate, m.p. 190° (MeOH),  $[\alpha]_D = 108^\circ$  (c 2·59; CHCl<sub>3</sub>); UV, IR and NMR spectra were identical with published data<sup>2,6</sup> (Found: C, 52·84; H, 5·88.  $C_{24}H_{32}O_{14}$  requires: C, 52·93; H, 5·93%).

Gentiopicroside tetraacetate was obtained by dehydration of 2 tetraacetate with  $Ac_2O-KHSO_4$ , m.p. 139–140° (MeOH),  $[\alpha]_D=180^\circ$  (c 0.86; CHCl<sub>3</sub>); UV and IR spectra were identical with the ones published.<sup>2,6</sup>

(b) Ethyl acetate extract. Evaporation of the EtOAc extract left 14 g of a crude brown residue which was chromatographed according to Koch et al.<sup>2</sup> over 130 g alumina. First fraction (Me<sub>2</sub>CO): 5·5 g, discarded; second fraction (Me<sub>2</sub>CO-H<sub>2</sub>O, 10:1) afforded 2·2 g (2), after acetylation (Ac<sub>2</sub>O-pyridine) gave 0·3 g of 2 tetraacetate; third fraction (Me<sub>2</sub>CO-H<sub>2</sub>O, 1:1) left 2·0 g (3), after acetylation (Ac<sub>2</sub>O-pyridine) 0·48 g of (+)-bornesitol pentaacetate were obtained, m.p. 145–146° (after softening above 140°) (MeOH),  $[\alpha]_D + 10^\circ$  (c 2·88; acetone), UV no absorption above 210 nm; MW 404 (ms), (Found: C, 50·21; H. 5·93; OCH<sub>3</sub>, 7·65. C<sub>16</sub>H<sub>21</sub>O<sub>10</sub>(Me)<sub>1</sub> requires: C, 50·35; H, 5·91; Mc, 7·63°6), <sup>1</sup>H-NMR: (a) 60 MHz, CDCl<sub>3</sub>,  $\delta$  2·02 (3 MeC=O), 2·06 (MeC=O), 2·19 (1 MeC=O), 3·36 (1 Me O), 3·53 (d, 1H), 4·85–5·85 (m, 4H) ppm (b) 100 MHz, CDCl<sub>3</sub>, Fourier Transform spectrum  $\delta$  4·95 (1H, I 10·0); I 2·28) 5·10 (1H, I 9·7) 5·35 (1H, I 10·0); 5·46 (1H, I 10·3), 5·72 (1H, I 2·275) ppm.

 $J_{a=a}10\cdot0$ ;  $J_{a=2}\cdot8$ ), 5·10 (1H,  $J_{u=a}9\cdot7$ ), 5·35 (1H,  $J_{a=a}10\cdot0$ ), 5·46 (1H,  $J_{a=a}10\cdot3$ ), 5·72 (1H,  $J_{a=2}\cdot75$ ) ppm. (+)-Bornesitol was obtained by hydrolysis of **2** pentaacetate with NH<sub>3</sub>-EtOH, m.p. 201–203°, [ $\alpha$ ]<sub>D</sub> + 29° ( $\alpha$ 3·80; H<sub>2</sub>O).

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<sup>4</sup> vide Comm. 2.

<sup>&</sup>lt;sup>6</sup> KUBOTA, T. and TOMITA, Y. (1961) Tetrahedron Letters, 453.